

Published on Web 12/13/2010

Core-Clickable PEG-*Branch*-Azide Bivalent-Bottle-Brush Polymers by ROMP: Grafting-Through and Clicking-To

Jeremiah A. Johnson,[†] Ying Y. Lu,[†] Alan O. Burts,[†] Yeon-Hee Lim,[‡] M. G. Finn,[‡] Jeffrey T. Koberstein,[§] Nicholas J. Turro,^{II} David A. Tirrell,^{*,†} and Robert H. Grubbs^{*,†}

Division of Chemistry and Chemical Engineering, California Institute of Technology, 1200 E. California Blvd., Pasadena, California 91125; Department of Chemistry and The Skaggs Institute for Chemical Biology, The Scripps Research Institute, 10550 North Torrey Pines Road, La Jolla, California 92037; Department of Chemical Engineering, Columbia University, 500 West 120th Street, New York, New York, 10027; and Department of Chemistry, Columbia University, 3000 Broadway, New York, New York 10027

Received September 18, 2010; E-mail: rhg@caltech.edu; tirrell@caltech.edu

Abstract: The combination of highly efficient polymerizations with modular "click" coupling reactions has enabled the synthesis of a wide variety of novel nanoscopic structures. Here we demonstrate the facile synthesis of a new class of clickable, branched nanostructures, polyethylene glycol (PEG)-*branch*-azide bivalent-brush polymers, facilitated by "graft-through" ring-opening metathesis polymerization of a branched norbornene-PEG-chloride macromonomer followed by halide-azide exchange. The resulting bivalent-brush polymers possess azide groups at the core near a polynorbornene backbone with PEG chains extended into solution; the structure resembles a unimolecular micelle. We demonstrate copper-catalyzed azide–alkyne cycloaddition (CuAAC) "click-to" coupling of a photocleavable doxorubicin (**DOX**)-alkyne derivative to the azide core. The CuAAC coupling was quantitative across a wide range of nanoscopic sizes (\sim 6– \sim 50 nm); UV photolysis of the resulting **DOX**-loaded materials yielded free **DOX** that was therapeutically effective against human cancer cells.

Introduction

The explosion of nanoscience is fuelled by problems that require nanoscale solutions and by the inherent challenges of nanoscale synthesis.^{1–3} In drug delivery, for example, plasma clearance time of a polymer–drug conjugate is closely linked to its nanoscopic size; materials smaller than 10 nm are excreted rapidly whereas larger materials often display significantly longer retention times.^{3–9} Nanoscopic architecture also plays an important role in determining in vivo retention time.^{10–12} Branched polymeric structures such as dendrimers, star poly-

- [‡] Department of Chemistry and The Skaggs Institute for Chemical Biology, The Scripps Research Institute.
 - [§] Department of Chemical Engineering, Columbia University.
 - ^{II} Department of Chemistry, Columbia University.
 - (1) Whitesides, G. M. Small 2005, 1, 172–179.
 - (2) Steigerwald, M. L.; Brus, L. E. Acc. Chem. Res. 1990, 23, 183-188.
 - (3) Peer, D.; Karp, J. M.; Hong, S.; Farokhzad, O. C.; Margalit, R.; Langer, R. *Nat. Nanotechnol.* **2007**, *2*, 751–760.
 - (4) Matsumura, Y.; Maeda, H. Cancer Res. 1986, 46, 6387-6392.
 - (5) Gref, R.; Minamitake, Y.; Peracchia, M. T.; Trubetskoy, V.; Torchilin, V.; Langer, R. Science 1994, 263, 1600–1603.
 - (6) Gref, R.; Domb, A.; Quellec, P.; Blunk, T.; Mueller, R. H.; Verbavatz, J. M.; Langer, R. Adv. Drug Delivery Rev. 1995, 16, 215–233.
 - (7) Edwards, D. A.; Hanes, J.; Caponetti, G.; Hrkach, J.; Ben-Jebria, A.; Eskew, M. L.; Mintzes, J.; Deaver, D.; Lotan, N.; Langer, R. *Science* **1997**, *276*, 1868–1871.
 - (8) Langer, R. Nature (London) 1998, 392, 5-10.
 - (9) Mammen, M.; Chio, S.-K.; Whitesides, G. M. Angew. Chem., Int. Ed. 1998, 37, 2755–2794.
- (10) Qiu, L. Y.; Bae, Y. H. Pharm. Res. 2006, 23, 1-30.

mers, bottle-brush polymers, and hyperbranched polymers generally display longer in vivo retention times compared to their linear polymer analogues.^{11,13} A wealth of research attention has focused on synthetic approaches to branched, stimuli-responsive, nanoscopic materials that possess orthogonal functionality for incorporation of therapeutic, imaging, and targeting molecules.^{14–25}

- (11) Fox, M. E.; Szoka, F. C.; Frechet, J. M. J. Acc. Chem. Res. 2009, 42, 1141–1151.
- (12) Chen, B.; Jerger, K.; Frechet, J. M. J.; Szoka, F. C. J. Controlled Release 2009, 140, 203–209.
- (13) Grayson, S. M.; Godbey, W. T. J. Drug Targeting 2008, 16, 329–356.
- (14) Duncan, R. Nat. Rev. Drug Discovery 2003, 2, 347-360.
- (15) Bartlett, D. W.; Su, H.; Hildebrandt, I. J.; Weber, W. A.; Davis, M. E. Proc. Natl. Acad. Sci. U.S.A. 2007, 104, 15549–15554.
- (16) Wu, P.; Malkoch, M.; Hunt, J. N.; Vestberg, R.; Kaltgrad, E.; Finn, M. G.; Fokin, V. V.; Sharpless, K. B.; Hawker, C. J. *Chem. Commun.* 2005, 5775–5777.
- (17) Ihre, H. R.; Padilla De Jesus, O. L.; Szoka, F. C., Jr.; Frechet, J. M. J. Bioconj. Chem. 2002, 13, 443–452.
- (18) Breitenkamp, K.; Emrick, T. J. Am. Chem. Soc. 2003, 125, 12070–12071.
- (19) Becker, M. L.; Remsen, E. E.; Pan, D.; Wooley, K. L. *Bioconj. Chem.* 2004, 15, 699–709.
- (20) Such, G. K.; Tjipto, E.; Postma, A.; Johnston, A. P. R.; Caruso, F. Nano Lett. 2007, 7, 1706–1710.
- (21) Cengelli, F.; Grzyb, J. A.; Montoro, A.; Hofmann, H.; Hanessian, S.; Juillerat-Jeanneret, L. *ChemMedChem* 2009, 4, 988–997.
- (22) Tong, G. J.; Hsiao, S. C.; Carrico, Z. M.; Francis, M. B. J. Am. Chem. Soc. 2009, 131, 11174–11178.
- (23) Lee, S.-M.; Chen, H.; O'Halloran, T. V.; Nguyen, S. T. J. Am. Chem. Soc. 2009, 131, 9311–9320.

 $^{^{\}dagger}$ Division of Chemistry and Chemical Engineering, California Institute of Technology.

The most widely studied branched macromolecules for drug delivery are dendrimers; their nanoscale, monodisperse, multivalent structures provide functional handles for elaboration with bioactive groups of interest.²⁶⁻²⁸ One factor that limits widespread application of dendrimers is synthetic difficulty. Though recent synthetic advances are impressive, 16,29-32 it remains a challenge to prepare functionally diverse dendrimers of variable sizes. To overcome this challenge, researchers have appended linear polymers to dendrimers^{33–36} or prepared linear polymers with dendritic fragments attached to their side chains. The latter "dendronized linear polymers" have attracted attention as nanoscopic building blocks; their cylindrical shapes can provide important advantages in nanoscale fabrication when compared to spherical dendrimers.³⁷⁻⁴³ Similar nanoscale structures can be formed from bottle-brush-polymers that carry long chains grafted at high density to linear polymer backbones.^{44–63} Bottle-

- (24) Lee, J.-S.; Green, J. J.; Love, K. T.; Sunshine, J.; Langer, R.; Anderson, D. G. Nano Lett. 2009, 9, 2402–2406.
- (25) Shen, Y.; Jin, E.; Zhang, B.; Murphy, C. J.; Sui, M.; Zhao, J.; Wang, J.; Tang, J.; Fan, M.; Van Kirk, E.; Murdoch, W. J. J. Am. Chem. Soc. 2010, 132, 4259–4265.
- (26) Helms, B.; Meijer, E. W. Science 2006, 313, 929-930.
- (27) Tomalia, D. A.; Baker, H.; Dewald, J.; Hall, M.; Kallos, G.; Martin, S.; Roeck, J.; Ryder, J.; Smith, P. *Polym. J.* **1985**, *17*, 117–132.
- (28) Bosman, A. W.; Janssen, H. M.; Meijer, E. W. Chem. Rev. 1999, 99, 1665–1688.
- (29) Montanez, M. I.; Campos, L. M.; Antoni, P.; Hed, Y.; Walter, M. V.; Krull, B. T.; Khan, A.; Hult, A.; Hawker, C. J.; Malkoch, M. *Macromolecules* **2010**, *43*, 6004–6013.
- (30) Antoni, P.; Robb, M. J.; Campos, L.; Montanez, M.; Hult, A.; Malmstrom, E.; Malkoch, M.; Hawker, C. J. *Macromolecules* 2010, *43*, 6625–6631.
- (31) Killops, K. L.; Campos, L. M.; Hawker, C. J. J. Am. Chem. Soc. 2008, 130, 5062–5064.
- (32) Wu, P.; Feldman, A. K.; Nugent, A. K.; Hawker, C. J.; Scheel, A.; Voit, B.; Pyun, J.; Frechet, J. M. J.; Sharpless, K. B.; Fokin, V. V. Angew. Chem., Int. Ed. 2004, 43, 3928–3932.
- (33) Lee, C. C.; Gillies, E. R.; Fox, M. E.; Guillaudeu, S. J.; Frechet, J. M. J.; Dy, E. E.; Szoka, F. C. *Proc. Natl. Acad. Sci. U.S.A.* 2006, *103*, 16649–16654.
- (34) Gillies, E. R.; Dy, E.; Frechet, J. M. J.; Szoka, F. C. Mol. Pharm. 2005, 2, 129–138.
- (35) Gillies, E. R.; Frechet, J. M. J. J. Org. Chem. 2004, 69, 46-53.
- (36) Gillies, E. R.; Frechet, J. M. J. J. Am. Chem. Soc. 2002, 124, 14137-
- 14146. (37) Grayson, S. M.; Frechet, J. M. J. *Macromolecules* **2001**, *34*, 6542–6544.
- (38) Boydston, A. J.; Holcombe, T. W.; Unruh, D. A.; Frechet, J. M. J.; Grubbs, R. H. J. Am. Chem. Soc. 2009, 131, 5388–5389.
- (39) Zhang, A.; Zhang, B.; Waechtersbach, E.; Schmidt, M.; Schlueter, A. D. Chem.-Eur. J. 2003, 9, 6083–6092.
- (40) Lee, C. C.; Grayson, S. M.; Frechet, J. M. J. J. Polym. Sci., Part A: Polym. Chem. 2004, 42, 3563–3578.
- (41) Yoshida, M.; Fresco, Z. M.; Ohnishi, S.; Frechet, J. M. J. Macromolecules 2005, 38, 334–344.
- (42) Canilho, N.; Kaseemi, E.; Mezzenga, R.; Schlueter, A. D. J. Am. Chem. Soc. 2006, 128, 13998–13999.
- (43) Helms, B.; Mynar, J. L.; Hawker, C. J.; Frechet, J. M. J. J. Am. Chem. Soc. 2004, 126, 15020–15021.
- (44) Zhang, M.; Mueller, A. H. E. J. Polym. Sci., Part A: Polym. Chem. 2005, 43, 3461–3481.
- (45) Jiang, X.; Lok, M. C.; Hennink, W. E. Bioconj. Chem. 2007, 18, 2077–2084.
- (46) Tsarevsky, N. V.; Bencherif, S. A.; Matyjaszewski, K. Macromolecules 2007, 40, 4439–4445.
- (47) Lutz, J.-F.; Boerner, H. G.; Weichenhan, K. Macromolecules 2006, 39, 6376–6383.
- (48) Allen, M. J.; Wangkanont, K.; Raines, R. T.; Kiessling, L. L. Macromolecules 2009, 42, 4023–4027.
- (49) Cheng, G.; Boeker, A.; Zhang, M.; Krausch, G.; Mueller, A. H. E. Macromolecules 2001, 34, 6883–6888.
- (50) Lu, H.; Wang, J.; Lin, Y.; Cheng, J. J. Am. Chem. Soc. 2009, 131, 13582–13583.
- (51) Morandi, G.; Pascual, S.; Montembault, V.; Legoupy, S.; Delorme, N.; Fontaine, L. *Macromolecules* 2009, 42, 6927–6931.

brush polymers are prepared by graft-to, graft-from, or graftthrough methodologies. The graft-to strategy requires an efficient coupling reaction to attach a functional molecule to every monomer unit of a linear polymer; "click"⁶⁴ reactions such as the copper-catalyzed azide—alkyne cycloaddition^{65,66} (CuAAC) and thiol—ene coupling have proven useful in this regard.^{43,46,67–70} Graft-from and graft-through methodologies require highly efficient polymerization reactions capable of initiation and propagation in sterically demanding environments.^{48,51–53,71} Recent developments in efficient, controlled polymerization coupled with new click reactions provide materials chemists with the tools to generate novel functional nanoscopic materials.^{72–77}

Ring-opening metathesis polymerization (ROMP) of strained alkene- (e.g., norbornene)-terminated macromonomers (MM) initiated by ruthenium *N*-heterocyclic carbene complexes (e.g., **1**, Figure 1) has recently proven useful for graft-through synthesis of functional bottle-brush polymers;^{50,78-83} fast initiation leads to low polydispersities (PDIs) while release of ring

- (52) Neugebauer, D.; Sumerlin, B. S.; Matyjaszewski, K.; Goodhart, B.; Sheiko, S. S. *Polymer* **2004**, *45*, 8173–8179.
- (53) Sumerlin, B. S.; Neugebauer, D.; Matyjaszewski, K. Macromolecules 2005, 38, 702–708.
- (54) Gao, H.; Matyjaszewski, K. J. Am. Chem. Soc. 2007, 129, 6633– 6639.
- (55) Hadjichristidis, N.; Pitsikalis, M.; Iatrou, H.; Pispas, S. Macromol. Rapid Commun. 2003, 24, 979–1013.
- (56) Tsukahara, Y.; Mizuno, K.; Segawa, A.; Yamashita, Y. Macromolecules 1989, 22, 1546–1552.
- (57) Tsukahara, Y.; Tsutsumi, K.; Yamashita, Y.; Shimada, S. Macromolecules 1990, 23, 5201–5208.
- (58) Dziezok, P.; Sheiko, S. S.; Fischer, K.; Schmidt, M.; Moller, M. Angew. Chem., Int. Ed. 1998, 36, 2812–2815.
- (59) Neiser, M. W.; Okuda, J.; Schmidt, M. *Macromolecules* 2003, 36, 5437–5439.
- (60) Neiser, M. W.; Muth, S.; Kolb, U.; Harris, J. R.; Okuda, J.; Schmidt, M. Angew. Chem., Int. Ed. 2004, 43, 3192–3195.
- (61) Yuan, Y.-Y.; Du, Q.; Wang, Y.-C.; Wang, J. *Macromolecules* **2010**, *43*, 1739–1746.
- (62) Li, C.; Ge, Z.; Fang, J.; Liu, S. *Macromolecules* **2009**, *42*, 2916–2924.
- (63) Li, A.; Lu, Z.; Zhou, Q.; Qiu, F.; Yang, Y. J. Polym. Sci., Part A: Polym. Chem. 2006, 44, 3942–3946.
- (64) Kolb, H. C.; Finn, M. G.; Sharpless, K. B. Angew. Chem., Int. Ed. 2001, 40, 2004–2021.
- (65) Tornoe, C. W.; Christensen, C.; Meldal, M. J. Org. Chem. 2002, 67, 3057–3064.
- (66) Rostovtsev, V. V.; Green, L. G.; Fokin, V. V.; Sharpless, K. B. Angew. Chem., Int. Ed. 2002, 41, 2596–2599.
- (67) Sumerlin, B. S.; Tsarevsky, N. V.; Louche, G.; Lee, R. Y.; Matyjaszewski, K. *Macromolecules* **2005**, *38*, 7540–7545.
- (68) Gao, H.; Matyjaszewski, K. J. Am. Chem. Soc. 2007, 129, 6633– 6639.
- (69) Tsarevsky, N. V.; Bernaerts, K. V.; Dufour, B.; Du Prez, F. E.; Matyjaszewski, K. Macromolecules 2004, 37, 9308–9313.
- (70) Campos, L. M.; Killops, K. L.; Sakai, R.; Paulusse, J. M. J.; Damiron, D.; Drockenmuller, E.; Messmore, B. W.; Hawker, C. J. *Macromolecules* **2008**, *41*, 7063–7070.
- (71) Boerner, H. G.; Beers, K.; Matyjaszewski, K.; Sheiko, S. S.; Moeller, M. *Macromolecules* **2001**, *34*, 4375–4383.
- (72) Golas, P. L.; Matyjaszewski, K. Chem. Soc. Rev. 2010, 39, 1338– 1354.
- (73) Iha, R. K.; Wooley, K. L.; Nystrom, A. M.; Burke, D. J.; Kade, M. J.; Hawker, C. J. Chem. Rev. 2009, 109, 5620–5686.
- (74) Hawker, C. J.; Wooley, K. L. Science 2005, 309, 1200-1205.
- (75) Johnson, J. A.; Finn, M. G.; Koberstein, J. T.; Turro, N. J. Macromol. Rapid Commun. 2008, 29, 1052–1072.
- (76) Binder, W. H.; Sachsenhofer, R. Macromol. Rapid Commun. 2007, 28, 15–54.
- (77) Hawker, C. J.; Fokin, V. V.; Finn, M. G.; Sharpless, K. B. Aust. J. Chem. 2007, 60, 381–383.
- (78) Xia, Y.; Kornfield, J. A.; Grubbs, R. H. *Macromolecules* **2009**, *42*, 3761–3766.
- (79) Xia, Y.; Olsen, B. D.; Kornfield, J. A.; Grubbs, R. H. J. Am. Chem. Soc. 2009, 131, 18525–18532.



Figure 1. Schematic depiction of the "graft-through and click-to" approach described in this work. A: We start with a bivalent macromonomer (2) and perform "graft-through" ROMP with catalyst 1 followed by in situ chloride-azide exchange. B: The resulting azido-bivalent-brush polymer is then functionalized with an alkyne by click chemistry in a "click-to" step. If the alkyne partner is a drug molecule linked via a cleavable linker, then the resulting brush polymer can release the drug in response to an external stimulus. In this work, we demonstrate controlled release of the anticancer agent doxorubicin (**DOX**) in response to 365 nm UV light.

strain provides the driving force for high conversion in these systems. Although catalyst 1 is incompatible with azides and alkynes, its functional group tolerance enables use of monomers bearing azide and alkyne precursors that are easily converted to the requisite click groups after polymerization. Binder and co-workers used ROMP combined with CuAAC to append various small molecules to clickable linear polymers.^{84,85} We have used ROMP in conjunction with CuAAC to generate cyclic polymers by end-to-end coupling.⁸⁶ To our knowledge, there are no examples of water-soluble brush polymers prepared by graft-through ROMP that are subsequently functionalized via click reactions. Such a strategy combines the benefits of ROMP for graft-through brush polymer synthesis with the modularity and efficiency of CuAAC for clicking-to; branched nanoscopic polymers with easily addressable functional groups can be rapidly prepared over a wide range of sizes simply by altering the ratio of catalyst to MM.

In this report, we demonstrate the sequential combination of graft-through- and click-to synthetic strategies to generate a

- (80) Li, Z.; Ma, J.; Cheng, C.; Zhang, K.; Wooley, K. L. Macromolecules 2010, 43, 1182–1184.
- (81) Li, Z.; Zhang, K.; Ma, J.; Cheng, C.; Wooley, K. L. J. Polym. Sci., Part A: Polym. Chem. 2009, 47, 5557–5563.
- (82) Conrad, R. M.; Grubbs, R. H. Angew. Chem., Int. Ed. 2009, 48, 8328– 8330.
- (83) Le, D.; Montembault, V.; Soutif, J. C.; Rutnakornpituk, M.; Fontaine, L. Macromolecules 2010, 43, 5611–5617.
- (84) Binder, W. H.; Kluger, C. *Macromolecules* 2004, *37*, 9321–9330.
 (85) Binder, W. H.; Kluger, C.; Josipovic, M.; Straif, C. J.; Friedbacher,
- G. Macromolecules 2006, 39, 8092–8101.
 (86) Clark, P. G.; Guidry, E. N.; Chan, W. Y.; Steinmetz, W. E.; Grubbs.
- (86) Clark, P. G.; Guidry, E. N.; Chan, W. Y.; Steinmetz, W. E.; Grubbs, R. H. J. Am. Chem. Soc. 2010, 132, 3405–3412.

clickable polyethylene glycol (PEG)-azide branched polymer material: a bivalent-bottle-brush polymer (Figure 1). Graftthrough ROMP of a PEG-chloride macromonomer (Figure 1, Scheme 1, 6) gave bivalent-brush polymers of varying nanoscopic sizes. These materials resemble first-generation dendronized polymers with branch points located near the linear polymer backbone (Figure 1); attached on one side of the branch point is a linear polymer and on the other side a small molecule (e.g., a drug). This design incorporates a water-soluble PEG domain that extends into solvent and a hydrophobic alkyl chloride near the core. We hypothesize that PEG attachment to one branch of these bivalent-brush polymers will confer the biocompatibility widely observed for PEGylated systems.^{6,87} Core-functionalized nanomaterials have been elaborated with functional small molecules by click chemistry;^{88–92} in this work, we convert the core alkyl chlorides to azides by treatment with sodium azide. Then, we utilize CuAAC to couple doxorubicin (DOX), to the core via a photocleavable nitrobenzyloxylcarbonyl (NBOC) linker. The latter enables controlled release of free DOX in response to long-wavelength (~365 nm) ultraviolet

- (87) Otsuka, H.; Nagasaki, Y.; Kataoka, K. Adv. Drug Delivery Rev. 2003, 55, 403–419.
- (88) O'Reilly, R. K.; Joralemon, M. J.; Hawker, C. J.; Wooley, K. L. New J. Chem. 2007, 31, 718–724.
- (89) O'Reilly, R. K.; Joralemon, M. J.; Hawker, C. J.; Wooley, K. L. *Chem.-Eur. J.* 2006, 12, 6776–6786.
- (90) O'Reilly, R. K.; Joralemon, M. J.; Wooley, K. L.; Hawker, C. J. *Chem. Mater.* 2005, 17, 5976–5988.
- (91) Joralemon, M. J.; O'Reilly, R. K.; Hawker, C. J.; Wooley, K. L. J. Am. Chem. Soc. 2005, 127, 16892–16899.
- (92) Prasuhn, D. E., Jr.; Singh, P.; Strable, E.; Brown, S.; Manchester, M.; Finn, M. G. J. Am. Chem. Soc. 2008, 130, 1328–1334.

Scheme 1. Synthesis of PEG-Norbornene-Chloride MM 6



(UV) light; we demonstrate that released **DOX** is therapeutically active against human cancer cells in culture.

Results and Discussion

Synthesis of MM 6. Clickable bivalent PEG brush polymers can be generated from an MM containing a strained norbornene derivative for ROMP, a PEG chain, and a functional group suitable for CuAAC (Figure 1, Scheme 1). Since neither azides nor alkynes are compatible with catalyst 1, we designed MM 6 to carry an alkyl chloride for facile conversion to an azide in a post-ROMP modification step. We incorporated a 6-carbon spacer between the strained norbornene moiety and the tertiary amide branch point to reduce steric hindrance between the growing polymer chain end and the branched side-chains. This spacer may also facilitate subsequent side chain modification reactions, such as chloride to azide conversion and CuAAC. MM 6 was obtained in five steps from *exo-N*-(6-hydroxyhexyl)-5-norbornene-2,3-dicarboximide (norb-C6OH, Scheme 1). The synthesis commenced with Swern oxidation of norb-C6OH to give aldehyde 2. Reductive amination using 2 and 3-chloropropylamine hydrochloride gave secondary amine 3, which was readily converted to acid 4 by treatment with succinic anhydride. Reaction of 4 with N-hydroxysuccinimide (NHS) in the presence of N-(3-dimethylaminopropyl)-N'-ethylcarbodiimide hydrochloride (EDC) and catalytic 4-(dimethylamino)pyridine (DMAP) gave NHS ester 5. We have prepared 5 on a multigram scale and expect it to be useful for the preparation of a variety of multifunctional ROMP monomers. In this work, commercially available 3 kDa PEG-NH₂ was treated with a slight excess of 5 to give MM 6. Excess 5 was used to ensure complete conversion of every PEG-amine to amide 6; the only polymeric species in the final reaction mixture was 6 and it was easily purified via repeated precipitation in diethyl ether. The ¹H NMR, ¹³C NMR, and MALDI spectra for MM 6 are provided in the Supporting Information.

Graft-Through ROMP of 6 and Halide-Azide Exchange to Generate nN_3 Bivalent-Brush Polymers. We pursued a one-pot synthetic strategy to rapidly generate a library of azide-functional brush polymers (nN_3) with variable number-average degrees of polymerization (DP) and hydrodynamic radii. A strength of this approach is the ease of generating branched functional materials **Scheme 2.** Synthesis of Azido-Bivalent-Brush Polymers (nN_3) by ROMP and Chloride-Azide Exchange



Table 1. Characterization of nN_3 Bivalent Brush Polymers by GPC (room temp., 0.2 M LiBr in DMF, 1 mL/min) and DLS (aqueous)

n (theo.) ^a	DP ^b	Mn (GPC, kDa)	PDI	<i>R</i> _h ∕∕nm
10	10	34.1	1.11	3.1 (0.3)
30	32	109	1.04	6.3 (0.8)
50	53	180	1.07	6.9 (0.9)
70	70	238	1.14	7.3 (0.3)
80	80	272	1.14	7.5 (0.8)
100	101	343	1.06	11(1)
300	295	1,000	1.11	15(1)
400	575	1,960	1.27	25 (2)

^{*a*} Ratio of MM to catalyst (i.e., theoretical DP). This number is used to identify \mathbf{nN}_3 and \mathbf{nDOX} samples throughout the text. ^{*b*} DP observed derived from $M_n(\text{GPC})/M_n(\text{MM})$. ^{*c*} Hydrodynamic radii measured by dynamic light scattering (DLS). DLS correlation functions were fit using the CONTIN algorithm.

of variable sizes rapidly and efficiently; each of the polymers described here was prepared by adding an aliquot of catalyst **1** to a vial containing MM **6**, allowing 60 min for ROMP to proceed, quenching the polymerization by addition of ethyl vinyl ether, solvent exchange, and finally treatment with sodium azide (Scheme 2). The first several steps were performed within a few hours, whereas the final step, attaching azides to the polymer side chains, was left for 48 h to ensure high conversion.

The polymer characterization data in Table 1 demonstrate that ROMP of **6** is well-controlled up to the highest DP tested; a small amount of catalyst deactivation and/or experimental error may explain the larger-than-expected DP for the **400N**₃ samples. In general, polymers with low polydispersities and molecular weights defined by the ratio of **6**:1 were obtained. The GPC traces for all samples (Figure 2A) show monomodal molecular weight distributions and very high (>95%) MM conversions. All of the polymers were highly soluble (>100 mg/mL) in water;



Figure 2. A: Representative GPC traces for nN_3 bivalent-brush polymer samples show monomodal molecular weight distributions, low polydispersities, and high conversions. B: Hydrodynamic diameter histograms for selected nN_3 polymers obtained from DLS measurements using the CONTIN fitting algorithm. The particle sizes increase with increasing DP and are varied over a wide range by adjusting the ratio of MM to catalyst (6:1) before graft-through ROMP.



Figure 3. FTIR spectra of nN_3 bivalent-brush polymers normalized to the strong PEG ether (C-O-C) antisymmetric stretch absorbance. The relative amount of azide is approximately the same for each n value which suggests that chloride-azide exchange is not sterically limited with increased DP.

aqueous DLS measurements (Table 1, Figure 2B) confirm that their nanoscopic dimensions increased with DP and were tunable over a broad range (radii from 3–25 nm) of relevant sizes for drug delivery applications. Furthermore, DLS (Figure 2B) shows that the particle size distributions are monomodal.

Graft-through ROMP ensures that every side chain unit possesses both a PEG chain and an alkyl chloride; the weight percentage of chloride is independent of DP. It follows that if every chloride is converted to an azide then the weight percentage of azide is also independent of DP. One may expect side chain modification reactions with bivalent-brush polymers to display DP-dependent conversion; perhaps increased DP would lead to lower reaction yields because of increased steric hindrance. We have found that for both chloride-azide exchange and CuAAC reactions there is no apparent difference in reaction yield across the DP values studied here. Figure 3 shows the FTIR spectra for a subset of nN_3 polymers normalized to the PEG ether (C-O-C antisymmetric stretch) absorbance at \sim 1100 cm⁻¹. The intensities of the characteristic azide antisymmetric stretch ($\sim 2100 \text{ cm}^{-1}$) are the same for each DP, which suggests that the azide:PEG ratio is the same for each sample and that the chloride-azide exchange reaction occurred with equal efficiency for each DP value.

Click Chemistry Using DOX-NBOC-Alkyne 9. The ultimate application of these nN_3 bivalent-brush polymers is defined by how they are decorated with functional groups. For drug delivery applications, one can imagine clicking a drug-alkyne derivative attached via a cleavable linker group (Figure 1). The resulting polymer would undergo controlled drug release in response to

an external stimulus leaving the brush polymer scaffold intact. In this work, we designed 9 for controlled release of **DOX** in response to UV light after click coupling to the nN_3 polymer scaffold (Scheme 3). Photoinitiated release from macromolecules provides a means for site-specific, controlled drug delivery and could potentially complement traditional phototherapeutic methods.^{93–103} Though placement of azide groups near the core of these bivalent-brush polymers should allow PEG to extend into solution and increase the biocompatibility of these materials, a buried location may make them difficult to functionalize with subsequent reactions. Anticipating this challenge was one reason we chose two highly efficient reactions, halide-azide exchange and CuAAC, as the modification reactions; as described above, CuAAC has proven useful for quantitative attachment of small molecules to the side chains of ROMP polymers^{84,85} and we expected that it would provide the best opportunity for highyield coupling of sterically demanding alkyne reagents to bivalent-branch polymers. There are a variety of CuAAC conditions throughout the literature and the catalyst/ligand/ solvent system depends on the application;^{104–107} perhaps the most impressive applications of CuAAC are in the field of

- (93) Dolmans, D. E. J. G. J.; Fukumura, D.; Jain, R. K. Nat. Rev. Cancer 2003, 3, 380–387.
- (94) Bethea, D.; Fullmer, B.; Syed, S.; Seltzer, G.; Tiano, J.; Rischko, C.; Gillespie, L.; Brown, D.; Gasparro, F. P. J. Dermatol. Sci. 1999, 19, 78–88.
- (95) Mal, N. K.; Fujiwara, M.; Tanaka, Y. Nature 2003, 421, 350-353.
- (96) Skwarczynski, M.; Noguchi, M.; Hirota, S.; Sohma, Y.; Kimura, T.; Hayashi, Y.; Kiso, Y. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 4492– 4496.
- (97) McCoy, C. P.; Rooney, C.; Edwards, C. R.; Jones, D. S.; Gorman, S. P. J. Am. Chem. Soc. 2007, 129, 9572–9573.
- (98) Jiang, M. Y.; Dolphin, D. J. Am. Chem. Soc. 2008, 130, 4236–4237.
 (99) Agasti, S. S.; Chompoosor, A.; You, C.-C.; Ghosh, P.; Kim, C. K.;
- Rotello, V. M. J. Am. Chem. Soc. 2009, 131, 5728–5729.
- (100) Lee, H.-i.; Wu, W.; Oh, J. K.; Mueller, L.; Sherwood, G.; Peteanu, L.; Kowalewski, T.; Matyjaszewski, K. Angew. Chem., Int. Ed. 2007, 46, 2453–2457.
- (101) Pastine, S. J.; Okawa, D.; Zettl, A.; Frechet, J. M. J. J. Am. Chem. Soc. 2009, 131, 13586–13587.
- (102) Kim, H.-C.; Hartner, S.; Behe, M.; Behr Thomas, M.; Hampp Norbert, A. J. Biomed. Opt. 2006, 11, 34024.
- (103) Haertner, S.; Kim, H.-C.; Hampp, N. J. Polym. Sci., Part A: Polym. Chem. 2007, 45, 2443–2452.
- (104) Lewis, W. G.; Magallon, F. G.; Fokin, V. V.; Finn, M. G. J. Am. Chem. Soc. 2004, 126, 9152–9153.
- (105) Rodionov, V. O.; Fokin, V. V.; Finn, M. G. Angew. Chem., Int. Ed. 2005, 44, 2210–2215.
- (106) Rodionov, V. O.; Presolski, S. I.; Diaz, D. D.; Fokin, V. V.; Finn, M. G. J. Am. Chem. Soc. 2007, 129, 12705–12712.
- (107) Rodionov, V. O.; Presolski, S. I.; Gardinier, S.; Lim, Y.-H.; Finn, M. G. J. Am. Chem. Soc. 2007, 129, 12696–12704.

Scheme 3. Synthesis of DOX-Nitrobenzyloxycarbonyl (NBOC)-Alkyne Derivative 9 for Click Chemistry and Photorelease of DOX



 $\begin{array}{l} \textbf{Scheme 4. } CuAAC \ Coupling \ of \ \textbf{9} \ to \ \textbf{nN}_3 \ Brush \ Using \\ Tris(hydroxypropyltriazolylmethyl)amine \ \textbf{(THPTA)} \ Ligand, \\ Copper(II) \ Sulfate, \ and \ Sodium \ Ascorbate \ As \ an \ in \ Situ \ Reductant \\ \end{array}$



bioconjugation where the requisite azide and alkyne functional groups are present in very low concentration among a number of other reactive groups.^{108–111} The Finn laboratory recently reported optimized CuAAC conditions for bioconjugation which utilize a new accelerating ligand THPTA (Scheme 4).¹¹² Aqueous CuAAC was found to proceed orders of magnitude faster with THPTA than in the absence of a ligand. Since these nN_3 bivalent-brush polymers are nanoscopic, narrowly dispersed, water-soluble materials with buried functional groups, we envisioned CuAAC reactions to them as similar to bioconjugation reactions to proteins bearing multiple azide or alkyne

(110) Speers, A. E.; Cravatt, B. F. Chem. Biol. 2004, 11, 535-546.

(112) Hong, V.; Presolski, S. I.; Ma, C.; Finn, M. G. Angew. Chem., Int. Ed. 2009, 48, 9879–9883. groups. After initial attempts without accelerating ligand, which led to <20% conversion after several days, we were pleased to find that addition of THPTA to the reaction mixture gave quantitative conversions. In a typical reaction, the polymer, THPTA (20 equiv. to azide), and alkyne **9** (1.05 equiv. to azide) were dissolved in 95% H₂O/5% DMSO such that the concentration of alkyne **9** was 2 mM. Aqueous sodium ascorbate (1.0 M) was added followed by copper(II) sulfate (1.0 M); two more aliquots of aqueous copper sulfate and sodium ascorbate were added over 2 days such that their final amounts were 10 and 50 equiv., respectively, relative to azide. The reactions were performed at 40 °C to achieve the maximum rate possible without inducing potential azide-olefin cycloaddition side reactions.⁸⁴

Figure 4A (inset) depicts the CuAAC coupling of 9 to 100N₃ as monitored by liquid-chromatography mass-spectrometry (LC-MS). Compound 9 eluted at ~4.2 min while DOX-loaded polymer eluted from \sim 3.5 - \sim 4 min. The unique absorbance of DOX (~500 nm) enables facile monitoring of the CuAAC reaction; the 100N₃ polymer does not absorb at 500 nm and therefore the growth of a new, broad polymer peak is attributed to CuAAC coupling of 9 to the polymer. The time-dependent LC-MS traces (Figure 4A, inset) show an increasing polymer absorbance at the expense of the alkyne 9 absorbance until a final, very high (>97%) conversion is reached. As for the case with chloride-azide exchange, one may expect the CuAAC reaction progress to depend on DP. Figure 4A shows preparatory high-performance liquid chromatography (prep-HPLC) 500 nm absorbance traces for several crude *n***DOX** polymers; the total ratio of *n*DOX:9 is the same for each DP which again suggests that DP has little effect on the efficiency of reactions to these structures. The retention times for the *n***DOX** polymers increase slightly for each DP indicating that polymer-column interactions are dependent on DP. The pure polymer fractions were collected and either lyophilized to dryness or concentrated before use in subsequent experiments. Figure 4B shows the FTIR spectrum of purified 50DOX compared to that of 50N₃. The azide antisymmetric stretch at $\sim 2100 \text{ cm}^{-1}$, present in the spectrum of 50N₃, is completely absent from the 50DOX spectrum which suggests very high consumption of the azide group. Integration of the broad aromatic resonances in the ¹H NMR spectrum of 50DOX compared to the PEG methylene groups is in close agreement to that expected for fully DOX-functionalized polymer (Figure S5). The final DOX weight percentage for the

⁽¹⁰⁸⁾ Ngo, J. T.; Champion, J. A.; Mahdavi, A.; Tanrikulu, I. C.; Beatty, K. E.; Connor, R. E.; Yoo, T. H.; Dieterich, D. C.; Schuman, E. M.; Tirrell, D. A. *Nat. Chem. Biol.* **2009**, *5*, 715–717.

⁽¹⁰⁹⁾ Dieterich, D. C.; Link, A. J.; Graumann, J.; Tirrell, D. A.; Schuman, E. M. Proc. Natl. Acad. Sci. U.S.A. 2006, 103, 9482–9487.

⁽¹¹¹⁾ Prescher, J. A.; Bertozzi, C. R. Nat. Chem. Biol. 2005, 1, 13-21.



Figure 4. A: Prep-HPLC traces of crude CuAAC mixtures which depict similar conversion for each *n* value. A (inset): Progress of CuAAC coupling of 9 to 100N₃ monitored by LC-MS. B: FTIR spectra of 50N₃ and 50DOX confirming complete loss of azide after CuAAC.



Figure 5. A: Time-dependent UV photolysis of **100DOX** monitored by LC–MS; after 10 min irradiation \sim 70% of free **DOX** was released. A minor product (labeled "*", see Scheme 5) was also observed at longer irradiation times. B: Viability of MCF-7 human breast cancer cells treated with free **DOX**, **100N**₃ bivalent-brush polymer, and drug-loaded **100DOX** polymer both with and without UV irradiation. Data points were fit to a sigmoidal function and the half-maximum inhibitory concentrations (IC₅₀) are shown for **DOX** and the photocleaved **100DOX**. The *x*-axis labels refer to the concentration of both free and polymer-conjugated drug.

functionalized polymers is 12.6%. As mentioned above, this value is independent of DP; it could be adjusted by varying the PEG length or the degree of branching in the MM precursor.

Photorelease of DOX and Cell Culture Studies. Having successfully coupled 9 to nN_3 we next studied the release of free DOX from the brush polymer in response to UV light (\sim 365 nm). An aqueous solution of purified *n***DOX** polymer was irradiated at 365 nm for a given time and subjected to LC-MS analysis. The progress of photorelease was monitored over 10 min by repeated irradiation and LC-MS; the data for **100DOX** are shown in Figure 5A. As irradiation time increased, we observed a new peak at ~ 3 min that increased in intensity at the expense of the 100DOX polymer peak. The new peak corresponds to the retention time and mass of free DOX. We also observed a small side product at \sim 3.8 min with a mass of 569 Da (labeled "*" in Figure 5A), which is likely the result of intramolecular trapping of carbamic acid 10 to give cyclic carbamate 11 (Scheme 5). Nevertheless, the major product of UV photolysis was free DOX; the photolysis yield after 10 min irradiation was \sim 70% for all *n***DOX** polymers which led us to examine the effectiveness of these materials against human cancer cells in cell culture.

We performed cell viability experiments using MCF-7 human breast cancer cells. Cells were treated with aqueous solutions of either free **DOX** or the corresponding *n***DOX** polymer at various concentrations and irradiated for 10 min using 365 nm light or kept in the dark. The cells were then incubated in the dark for 24 h, washed twice, and incubated for another 24 h in fresh, drug-free growth medium. Cell viability was assessed using the MTT assay (see Methods and Materials for details). Data for n = 100 polymers (100N₃ and 100DOX) are shown in Figure 5B (cell viability data for other n values is given in the Supporting Information). Free DOX, with and without UV irradiation, effectively killed the cells at $3-5 \mu M$ while azide polymer 100N₃ was nontoxic with and without light up to 100 μ M; these data taken together suggest that under our conditions 10 min of 365 nm UV irradiation has no adverse affect on cell viability nor does it interrupt the cellular toxicity of DOX. None of the *n***DOX** bivalent-brush polymers studied were toxic up to 50 μ M in the absence of UV irradiation; irradiation for 10 min yielded IC₅₀ values $\sim 7-10 \,\mu$ M which confirms that free **DOX** generated by photorelease is therapeutically active. The IC_{50} values were not DP-dependent. This observation suggests that the *n***DOX** polymers remain in the extracellular environment

Scheme 5. Photolysis of nDOX Polymers Gives Carbamic Acid 10ª



^{*a*} Decarboxylation of **10** yields free **DOX** as the major product while intramolecular trapping provides **DOX**-carbamate derivative **11** as a minor product. Reactive nitrosobenzaldehyde photoproducts remain bound to the polymer core after photolysis.

and upon photolysis free **DOX** is released and diffuses into the cell nucleus where it induces apoptosis. Although in vitro experiments do not show DP-dependent cytotoxicity, we expect that DP will affect in vivo trafficking and thus the overall utility of these materials in drug delivery. Targeting groups could be appended to the periphery of these materials to promote trafficking into the cell.¹¹³

Conclusions

Here we have demonstrated the utility of a combined graftthrough/click-to strategy for the synthesis of a new class of clickable, branched nanostructures: PEG-*branch*-azide bivalent-brush polymers. This approach makes use of the remarkable efficiency of ROMP for graft-through polymerization and the modular coupling power of click chemistry for polymer functionalization. A wide range of nanoscale sizes is accessible which will enable detailed structure/function correlation in biological settings. We are currently exploring applications for these materials in drug delivery. Design and synthesis of novel clickable linkers (e.g., longer wavelength-,

(113) Kolonko, E. M.; Kiessling, L. L. J. Am. Chem. Soc. 2008, 130, 5626– 5627. pH-, and redox-cleavable), attachment of targeting moieties at the PEG periphery, incorporation of degradable units within the polymer backbone, and use of chain-transfer agents to append orthogonal groups selectively to the ends of these polymers are being studied. Furthermore, copolymerization of MMs like **6** with other monomers (such as a norbornene-alkyl halide small molecule) will provide access to new multiblock polymer structures with the potential for higher drug loadings.

Acknowledgment. We thank Dr. S. Virgil and Mr. S. Presolski for helpful advice. This work was supported by the National Institutes of Health (NIH, R01-GM31332), the Beckman Institute at Caltech (postdoctoral fellowship for J.A.J.), and the MRSEC program of the National Science Foundation (NSF) under Award No. DMR-0520565.

Supporting Information Available: Synthetic procedures, spectral data, liquid chromatography methods, and details of cell viability studies. This material is available free of charge via the Internet at http://pubs.acs.org.

JA108441D